
Application of Artificial Neural Networks in Classification of Autism Diagnosis Based on Gene Expression Signatures

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6 **Abstract**

7 Clinical genetic tests of autism spectrum disorder (ASD) have largely been
8 difficult to develop due to the complex and heterogeneous nature of
9 autism's dependence on genetic factors. No single genetic component
10 accounts for all cases of autism, and large genetic variation is observed
11 across individuals. Thus, genetic data must be analyzed on a genomic scale
12 in the creation of a generalized autism diagnosis method based on genetics.
13 Artificial neural networks, when presented with enough examples, can learn
14 to correctly classify large sets of input. This project utilizes multilayer
15 perceptron network architecture with a backpropagation training method in
16 order to develop a trained network that can distinguish toddlers as having
17 ASD or not when fed large microarray genetic expression data. Successful
18 identification of ASD toddlers was able to be accomplished at a 73% rate.

19
20 **1 Introduction**

21 Long term prognosis of individuals with autism spectrum disorder (ASD) can improve
22 dramatically with early intervention [1][2], thus making early detection critical to the
23 treatment of ASD. Unfortunately, the current most reliable test for early autism diagnosis,
24 the Autism Diagnostic Observation Schedule-Toddler Module [3], is based on behavioral
25 observations, which limits age of detection to when clinically defined behavioral symptoms
26 begin to manifest. Rather than relying on behavioral data, diagnostic approaches based on
27 genetics may further decrease the age at which ASD can be detected. Numerous studies have
28 indicated that autism has a strong genetic basis, although the genetic mechanisms that lead to
29 the highly variable ASD phenotypes are complex and poorly understood.

30 Supervised machine learning, such as artificial neural networks (ANN), has been
31 successfully implemented to predict cancer classifications that typically elude routine
32 histology [4] [5]. The goal of this project is to train an ANN to return output of correct ASD
33 diagnoses when given input microarray data collected from toddlers with ASD or typically
34 developing (TD) toddlers. The ANN in this project will be built using Matlab's Neural
35 Network Toolbox [6] with the specific approach of applying the feedforward resilient
36 backpropagation learning algorithm to a multilayer perceptron. Multilayer perceptrons were
37 chosen on the assumption that subjects with ASD and those without ASD are not linearly
38 separable. A major goal of this project is to minimize the size of the input set for optimal
39 classification performance.

40
41 **2 Methods**

42 **2.1 Microarray Input**

43 Microarray genome-wide gene expression levels were obtained using the Illumina Humanht
44 12 v4 Expression BeadChip (Illumina, San Diego, CA, USA). Gene expression levels were
45 collected from 222 toddlers (137 ASD, 85 TD) aged 18-36 months. Of the 222 subjects, 150
46 were used for training and 72 were used for independent testing. Differential expression
47 levels were logged to give equal weight to ratios less than 1 and ratios greater than one. 835
48 differentially expressed genes were identified. To reduce the dimensionality of the data,
49 principal component analysis was done and the top 10 principal components were selected
50 such that at least 80% of the variance is captured. These data manipulations resulted in a x
51 10 matrix that can then be fed into the multilayer perceptron for training.

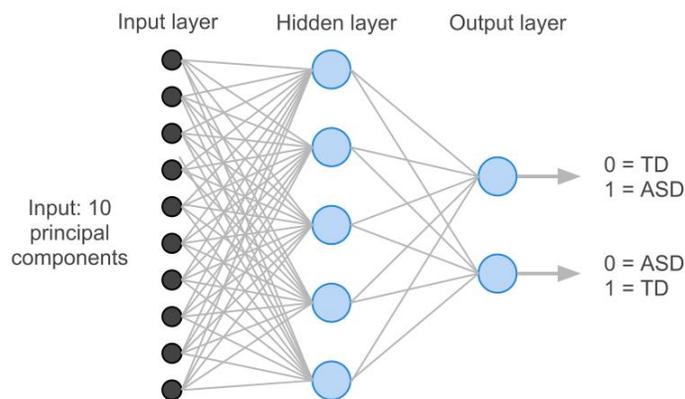
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53 **2.2 Cross-Validation and Committee Voting**

54 As part of the training process, a portion of the training set is set aside for validation such
55 that the network will know when to stop training to prevent overtraining and maintain ability
56 to generalize. Training is stopped when performance on the validation set fails to improve
57 after 6 iterations of training. 10-fold cross-validation groups were randomly selected such
58 that 1 group will be used for validation and the other 9 groups will be used for learning.
59 Subsequent networks will cycle through each of the 10 cross-validation groups to use as the
60 validation set. Thus, 10 networks will be trained per instance of cross-validation. After
61 training, an independent sample of 72 test subjects was fed into the network to test the
62 network. To prevent effects of uneven random partitioning of data, cross-validation will be
63 repeated many times and the outputs from networks will be averaged across these groups to
64 determine a committee vote for classification. A committee size of 100 was used, for a total
65 of 1000 networks trained per committee vote.

66
67 **2.3 Network Architecture**

68 Networks with multiple layers are capable of solving non-linearly separable classification
69 problems. The feedforward multilayer perceptron architecture used in this project only had
70 one hidden layer between the input and output layer, for a total of 2 layers of neurons
71 (Figure 1). The first layer had 5 neurons for receiving input, and the second layer had 2
72 neurons for producing output. One output neuron classifies TD subjects as 0 and ASD
73 subjects as 1, while the other neuron classifies ASD subjects as 0 and TD subjects as 1.
74 Even the ASD/TD classification is 2-bit information and can be explained with only one
75 neuron, 2 output neurons were used to observe training discrepancies between the 2 output
76 neurons, since they did not always perform equally.

77
78



79 **Figure 1: Network architecture**

80

81 Information flow from input to output first occurs by multiplying each input into each

82 neuron of the hidden layer by a weight value. The output (a_1) of the hidden layer is
 83 calculated by multiplying a matrix of weights (W_1) from each input to each neuron in the
 84 hidden layer by the matrix of input values (p) for one subject from plus a matrix of bias
 85 values (b_1) for each neuron in the hidden layer:

86

$$a_1 = \text{tansig}(W_1 p + b_1),$$

$$\text{tansig}(n) = \frac{e^n - e^{-n}}{e^n + e^{-n}}$$

87

88 The output (a_2) of the output layer is calculated by multiplying a matrix of weights (W_2)
 89 from each output of the hidden layer to each neuron in the output layer by the matrix of
 90 output values from the hidden layer (a_1) for one subject from plus a matrix of bias values
 91 (b_2) for each neuron in the output layer:

92

$$a_2 = \text{purelin}(W_2 a_1 + b_2),$$

$$\text{purelin}(n) = n$$

93

94

95 The transfer function *tansig* was chosen for the hidden layer because it is differentiable and
 96 has inputs and outputs that can be any real number.

97

98 **2.4 Training**

99

100 Each network will be trained using the resilient backpropagation algorithm [7][8] in the
 101 Matlab Neural Network Toolbox. Backpropagation is a method of updating weights and
 102 biases after each example of input is fed into the network. First, the approximate
 103 performance index is calculated as:

104

$$105 \quad \hat{F}(x) = e^T(k) e(k) = (t(k) - a(k))^T.$$

106

107 where t is the target value, a is the final output value, k is the input expression value for one
 108 gene, and T is a superscript indicating the connection.

109 The derivative of the performance index is used to calculate the sensitivity of the
 110 performance index to changes in each element of the net input (n^m) at neuron layer m .

$$s^m = \frac{\partial \hat{F}}{\partial n^m}$$

111

$$s^M = -2\hat{F}^M(n^M)(t - a)$$

112

$$113 \quad s^m = \hat{F}^m(n^m)(W^{m+1})^T s^{m+1}, \text{ for } m = M - 1, \dots, 2, 1.$$

114

115 The term “resilient” in resilient backpropagation refers to the method for updating weights
 116 and biases. The sigmoid transfer function used in this network is characterized by the fact
 117 that their slope must approach zero as the input becomes large. This results in a gradient that
 118 can have a very small magnitude and thus result in small changes in weights and biases. To
 119 circumvent these problems, only the sign, but not the magnitude, of the derivative is used to
 120 determine a weight change. Each weight and bias update values increases by a factor of a

121 specified $\delta_{increase}$ whenever the derivative of the performance function with respect to that
 122 weight has the same sign for 2 successive iterations. Similarly, each weight and bias update
 123 values decreases by a factor of a specified $\delta_{decrease}$ whenever the derivative of the
 124 performance function with respect to that weight changes sign from the previous iteration.
 125 For the training of the networks in this project, $\delta_0 = 0.7$, $\delta_{increase} = 1.2$ and $\delta_{decrease} = 0.5$,
 126 and $\delta_{max} = 50$.

127

128 2.5 Gene Minimization

129 It was unknown a priori how many and which of the total 835 genes were relevant to ASD
 130 diagnosis, so the initial dataset was noisy. To reduce the noise of the dataset, a gene
 131 minimization algorithm [7] was implemented after training 1000 networks (10-fold cross-
 132 validation x 100 committee size) with the full 835 genes. Gene minimization entailed
 133 ranking each gene based on the output's sensitivity to each gene. Sensitivity (S) of output (o)
 134 with respect to each of the 835 gene variables (x_k) were calculated as:

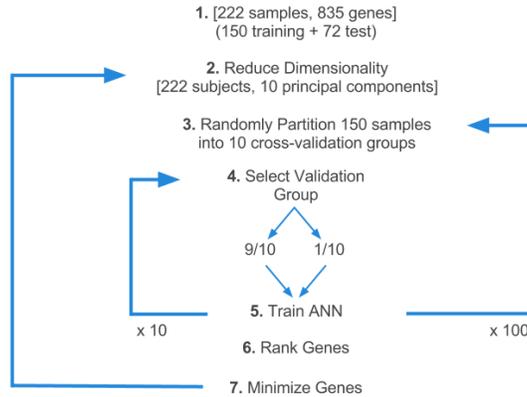
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$$S_k = \frac{1}{N_s} \frac{1}{N_o} \sum_{s=1}^{N_s} \sum_{i=1}^{N_o} \left| \frac{\partial o_i}{\partial x_k} \right|$$

136

137 where N_s is the number of subjects (150) and N_o is the number of outputs (2). Genes were
 138 ranked based on their sensitivity values, and then a subset of the highest ranked genes was
 139 used to train a new committee of networks.

140



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Figure 2: Methods overview

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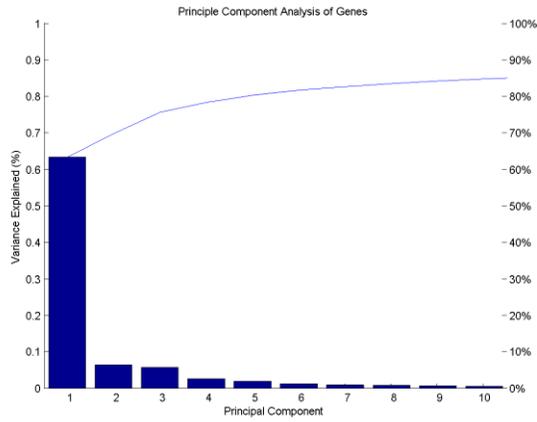
144 3 Results

145

146 3.1 Principal Component Analysis

147 Using principal component analysis, the 835 genes were reduced to 10 principal components
 148 that collectively captured 84.64% of the variance (Figure 3).

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Figure 3: Principal component analysis

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3.2 Training with 835 Genes

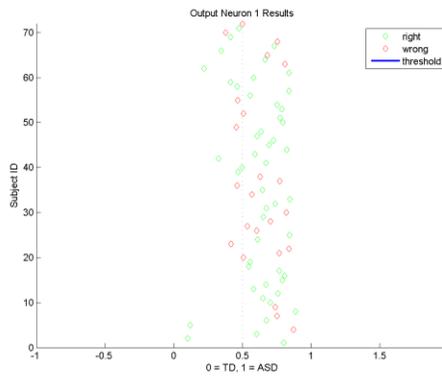
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Training with 835 genes produced a committee voting that produced 68.0556% correct output from Output Neuron 1 (Figure 4) and 68.0556% correct output from Output Neuron 2 (Figure 5).

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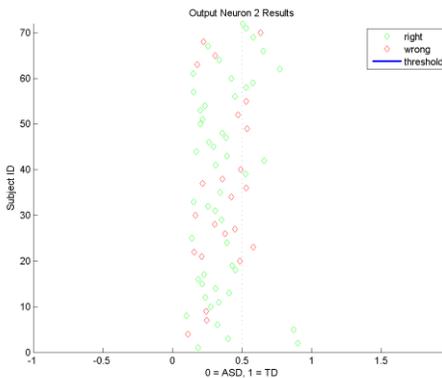
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Figure 4: Output Neuron 1 results using 835 genes



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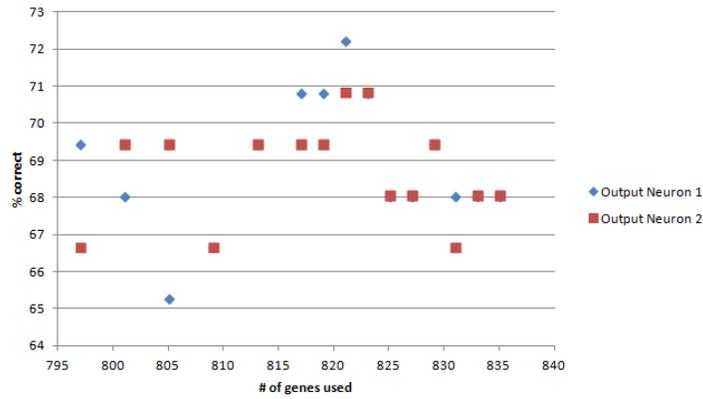
Figure 5: Output Neuron 2 results using 835 genes

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161 **3.2 Gene Minimization**

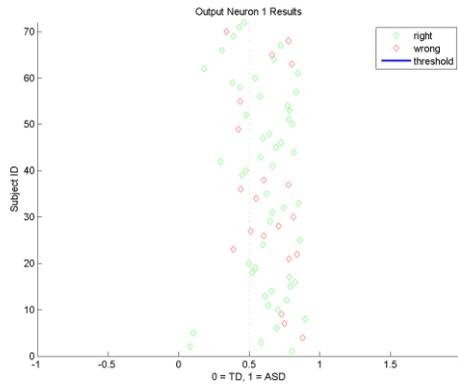
162 Sensitivities were calculated for each gene, and various subsets of the highest ranking genes
163 were used for training the networks. Performance peaked when 821 genes were used and
164 declined quickly with decreasing number of genes (Figure 6). Training with 821 genes
165 produced a committee voting that produced 72.2222% correct output from Output Neuron 1
166 (Figure 7) and 70.8333% correct output from Output Neuron 2 (Figure 8).

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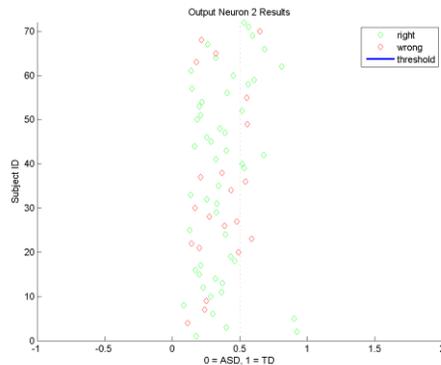
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Figure 6: Gene Minimization



172

Figure 7: Output Neuron 1 results using 821 genes



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174

Figure 8: Output Neuron 2 results using 821 genes

175 **4 Discussion**

176 While this project's artificial neural network architecture was able to increase classification
177 above random chance (50%), the highest success rate achieved (72.222%) is not high enough
178 to be of substantial clinical application. Even though some improvement (4.1666%) in
179 correct classification was obtained through gene minimization, the gain was not substantial.
180 The likely problem with the classification problem is that ASD is a very heterogeneous
181 disorder that may have subgroups with drastically different genetic expression signatures. To
182 improve classification, it may be useful to stratify the ASD class into subgroups and enrich
183 the input set with clinical measures.

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185 **References**

- 186 [1] Dawson, G., & Osterling, J. (1997). "Early intervention in autism." The effectiveness of early
187 intervention, 307-326.
- 188
- 189 [2] Rogers, S. J. (1998). "Empirically supported comprehensive treatments for young children with
190 autism." *Journal of Clinical Child Psychology*, 27(2), 168-179.
- 191
- 192 [3] Luyster, R., Guthrie, W., Gotham, K., Risi, S., DiLavore, P., & Lord, C. (2009). "The Autism
193 Diagnostic Observation Schedule-Toddler module: Preliminary findings using a modified version of
194 the ADOS."
- 195
- 196 [4] Greer, B. T., & Khan, J. (2004). "Diagnostic classification of cancer using DNA microarrays and
197 artificial intelligence." *Annals of the New York Academy of Sciences*, 1020(1), 49-66.
- 198
- 199 [5] Greer, B., & Khan, J. (2007). Online analysis of microarray data using artificial neural
200 networks. *METHODS IN MOLECULAR BIOLOGY-CLIFTON THEN TOTOWA-*, 377, 61
- 201
- 202 [6] Demuth, H., & Beale, M. (1993). *Neural network toolbox for use with MATLAB*.
- 203
- 204 [7] Khan, J., Wei, J. S., Ringner, M., et al. (2001) Classification and diagnostic prediction of cancers using
205 gene expression profiling and artificial neural networks. *Nat.Med.* 7, 673-679
- 206
- 207 [8] Demuth, H., Beale, M., & Hagan, M. (1992). *Neural Network Toolbox™ 6. User Guide,*
208 *COPYRIGHT, 2008.*